Tetrahedron 66 (2010) 6383–6390

Contents lists available at ScienceDirect

Tetrahedron

journal homepage: www.elsevier.com/locate/tet

Dihydro-3-(triphenylphosphoranylidene)-2,5-thiophendione: a convenient synthon for the preparation of substituted 1,4-thiazepin-5-ones and piperidinones via the intermediacy of thioacids

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ABSTRACT

article info

Article history: Received 20 December 2009 Received in revised form 27 March 2010 Accepted 1 April 2010 Available online 8 April 2010

Keywords: Thioacid Amide Coupling Thioanhydride Conjugate addition Radical reactions

Reaction of thiomaleic anhydride with triphenylphosphine gives the title compound, which undergoes reaction with a variety of aldehydes to give a range of alkylidene thiomaleic anhydrides (substituted monothioitaconic anhydrides). Subsequent treatment with tert-butoxycarbonylamino-substituted thiols, or under radical conditions with tert-butoxycarbonylamino-substituted alkyl halides results in a series of substituted monothiomaleic anhydrides, that on exposure to trifluoroacetic acid and then base lead to thiocarboxyl substituted 1,4-thiazepin-5-ones and piperidinones, respectively, that are ultimately trapped by reaction with 2,4-dinitrobenzenesulfonamides to give the corresponding amides.

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1. Introduction

Monothioacids^{1–6} present numerous advantages over the simple carboxylic acids by virtue of their acidity and the enhanced nucleophilicity of the thiocarboxylate function. Thus, for example, thioacids may be alkylated selectively in the presence of acids, $⁷$ $⁷$ $⁷$ and</sup> react with electrophiles such as electron-deficient aromatic and heteroaromatic systems, $^{8-11}$ isothiocyanates, and isocyanates, 12,13 isonitriles, $^{14-17}$ a variety of coupling reagents, $^{18-21}$ azides, $^{22-30}$ and alkenes^{[31–34](#page-7-0)} more efficiently and under milder conditions than the corresponding acids. In this context we^{[35,36](#page-7-0)} and others^{[37](#page-7-0)} have been exploring new methods for the preparation of a variety of thioacids for subsequent application in amide and peptide bond formation. In particular we have been interested in methods based on the nucleophilic ring opening of monothio thiosuccinic and thiomaleic anhydrides, $38-42$ and of monothio- β -lactones^{[43](#page-7-0)} and on the application of the ensuing thioacids in multicomponent coupling processes. Here, we report on the preparation of dihydro-3-(triphenylphosphoranylidene)-2,5-thiophendione, its elaboration to a variety of monothioitaconic anhydride derivatives, and the use of these latter products in multicomponent coupling reactions leading to the formation of 1,4-thiazepin5-ones.

2. Results and discussion

By analogy with the chemistry of 2-(triphenylphosphanylidenesuccinic anhydride), $44-51$ we reasoned that treatment of mono-thiomaleic anhydride^{[39,40,52,53](#page-7-0)} with triphenylphosphine would result in the formation of dihydro-3-(triphenylphosphoranylidene)-2,5 thiophendione 1, and that this ylide would be stable, isolable, and a suitable partner for reaction with aldehydes to give substituted monothioitaconic anhydride derivatives. In the event, reaction of monothiomaleic anhydride with triphenylphosphine in glacial acetic acid at room temperature provided the crystalline 1 in 86% yield (Scheme 1).

Scheme 1. Formation of dihydro-3-(triphenylphosphoranylidene)-2,5-thiophendione.

Subsequent reaction with a selection of aromatic and aliphatic aldehydes at room temperature gave the corresponding substituted monothioitaconic anhydrides in excellent yields as the pure E-isomers ([Table 1\)](#page-1-0), as ascertained by NOE studies, in agreement with the comparable reactions⁴⁶ of 2-(triphenylphosphanylidene)succinic anhydride.

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^{0040-4020/\$ –} see front matter © 2010 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2010.04.002

Table 1

Olefination reactions of reagent 1

With a series of alkylidene monothiosuccinic anhydrides in hand, we turned our attention to tandem Michael addition–nucleophilic ring opening processes with suitably constituted mercaptoamines, of the kind previously practiced on monothiomaleic anhydride, [39,40](#page-7-0) with subsequent trapping of the released thioacid by reaction with 2,4-dinitrobenzenesulfonamides. $9-11,36$ A series of reactions were conducted in which either 2-mercaptoaniline or 2-mercaptoethylamine were stirred in DMF at 0° C overnight with the alkylidene thiosuccinic anhydride before the addition of cesium carbonate and a 2,4-dinitrobenzenesulfonamide leading, ultimately, to the isolation of a series of substituted 1,4-thiazepin-5-ones, or their benzo-fused analogs, as presented in [Table 2.](#page-2-0)

For each of the tandem sequences reported in [Table 2](#page-2-0) only the thiazepinone regioisomer, arising from ring closure of the amine onto the proximal carbonyl group following conjugate addition, was iso-lated.⁵⁴ In the case of the ethylidene succinic anhydride [\(Table 2,](#page-2-0) entries 5–8), two diastereomeric products were formed in an approximate equimolar ratio and no attempt was made to separate the two isomers. With moderately more bulky octylidene and cyclopropylidene systems 4 and 5 ([Table 2,](#page-2-0) entries 9–12), low to moderate selectivity was observed in favor of the trans-isomer, whereas in the case of the somewhat more hindered benzylidene system 2 the transisomer was typically favored by a factor of 4–5:1 over the cis-isomer ([Table 2,](#page-2-0) entries 1–4). This preferential formation of the trans-isomers, with selectivity increasing with the size of the olefinic substituent, is best rationalized in terms the standard models for stereoselective protonation of enolates bearing a stereogenic center β -to the incipient carbonyl group.⁵⁵⁻⁶⁴ Thus, following conjugate addition of the thiolate anion, the enolate adopts a conformation that minimizes $A^{1,3}$ strain^{65,66} leading to protonation through either of transitions states A or B [\(Scheme 2\)](#page-3-0). As the size of the alkene substitutent (R) increases transition state **B** is increasingly favored, resulting in overall increased trans-selectivity.⁶⁷

Having demonstrated the 1,4-addition sequence with aminothiols, we turned our attention to the conjugate addition of functionalized carbon radicals and the preparation of piperidinone-based systems. Thus, radical addition of N-(tert-butoxycarbonyl) 2-iodoethylamine **22** to the ethylidene succinic anhydride 3 in toluene at 80 $^{\circ}$ C under the aegis of tris(trimethylsilyl)silane (TTMS)⁶⁸ and azobisisobutyronitrile (AIBN), gave the adduct 23 in 52% yields as a 3:1 anti/syn mixture of diastereoisomers [\(Scheme 3\)](#page-3-0), whose stereochemistry was assigned following subsequent conversion to the cis- and trans-piperidinones, respectively (vide infra). The stereoselectivity of this radical addition is rationalized by a model directly analogous to that advanced for enolate protonation in [Scheme 2.](#page-3-0) Thus, hydrogen atom transfer to the a-carbonyl radical occurs from the face opposite the more bulky alkyl chain in a conformation that minimizes $A^{1,3}$ strain, as is the case for other related diastereoselective radical reactions.⁶⁹⁻⁷⁴

Subsequent treatment of this mixture of diastereomers with trifluoroacetic acid removed the tert-butyloxycarbonyl protecting groups resulting, after removal of the acid under vacuum and treatment with base, in the cyclization of the liberated amine onto the proximal carbonyl carbon and the formation of the piperidinone skeleton bearing a thioacid substituent. Without isolation, this last species was captured by reaction with electron-deficient sulfonamides resulting overall in the piperidinonyl acetamides 24 and 25 ([Scheme 4](#page-4-0)). $54,75$

3. Conclusion

Dihydro-3-(triphenylphosphoranylidene)-2,5-thiophendione is readily accessed by reaction of monothiomaleic anhydride with triphenylphosphine. It reacts with a range of aldehydes in highly E-selective Wittig reactions to give substituted monothioitaconic anhydride derivatives. These latter substances undergo conjugate addition reactions with tert-butoxycarbonylamino-derived thiols and alkyl radicals to give a series of adducts that, on treatment with trifluoroacetic anhydride followed by base, lead to thiocarboxylsubstituted heterocycles. Finally, the thiocarboxylates can be trapped in situ by reaction with 2,4-dinitrobenzenesulfonamides to give the corresponding amides.

4. Experimental section

4.1. Dihydro-3-(triphenylphosphoranylidene)-2,5 thiophendione (1)

Triphenylphosphine (2.62 g, 10 mmol) was added to monothiomaleic anhydride (1.14 g, 10 mmol) in glacial acetic acid (5 mL) at room temperature and stirred for 3 h. Removal of the solvent under vacuum at room temperature and trituration of the residue with diethyl ether (3×5 mL) afforded an off-white solid in 86% yield. mp: 137–137.5 °C (decomp.); IR (film) 1700 and 1611 cm⁻¹; ¹H NMR $(500$ MHz, CDCl₃) δ 7.69-7.67 (m, 9H), 7.56 (dt, J=2.5, 7.5 Hz, 6H), 3.34 $(d, J=1 Hz, 2H);$ 13C NMR (125 MHz, CDCl₃) δ 205.6,178.8,133.6,129.6, 124.8,124.07, 50.0; ³¹P NMR (400 MHz, CDCl₃) δ 12.78; ESIHRMS, m/z calcd for C₂₂H₁₇O₂PSNa (M+Na)⁺: 399.0585, found: 399.0588.

4.2. General procedure for Wittig reactions

Reagent 1 (376 mg, 1 mmol) was stirred with excess aldehyde (1 mL) at room temperature for 10 h. Excess aldehyde was removed under vacuum at room temperature and the residue was purified by chromatographic purification over silica gel, that had been prewashed with acetone followed by hexanes, to give the alkylidene monothiosuccinic anhydrides.

4.2.1. E-2-(Benzylidene)thiosuccinic anhydride (2). Rapid chromatographic purification over silica gel eluting with 10% EtOAc in hexanes afforded a pale yellow oil in 78% yield. IR (film) 1731 and 1692 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.68 (t, J=2.5 Hz, 1H),

Table 2

Reaction of alkylidene monothiosuccinic anhydrides with mercaptoamines and 2,4-dinitrobenzenesulfonamides

1.
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$$

Table 2 (continued)

Scheme 2. Model for stereoselective protonation en route to the 1,4-thiazepin-5-ones.

Scheme 3. Radical addition to ethylidene monothiosuccinic anhydride.

7.51–7.46 (m, 5H), 3.98 (d, J=2.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) d 197.7, 192.7, 135.3, 133.8, 131.2, 131.1, 130.8, 130.1, 129.5, 128.4, 45.3; ESIHRMS m/z calcd for C₁₁H₇O₂S (M-1)⁻: 203.0172, found: 203.0180.

4.2.2. E-2-(Ethylidene)thiosuccinic anhydride (3). Rapid chromatographic purification over silica gel eluting with 10% EtOAc in hexanes afforded a colorless oil in 76% yield. IR (film) 1742 and 1706 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 6.99-6.94 (m, 1H), 3.66-3.65 (m, 2H), 1.91 (dt J=2.0, 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 197.6, 191.3, 135.2, 134.0, 43.2, 16.5; ESIHRMS m/z calcd for $C_6H_5O_2S (M-1)^{-}$: 141.0016, found: 141.0022.

4.2.3. E-2-(Octylidene)thiosuccinic anhydride (4). Rapid chromatographic purification over silica gel eluting with 10% EtOAc in hexanes afforded a colorless oil in 72% yield. IR (film) 1742 and 1706 $\rm cm^{-1}$; $\rm ^1H$ NMR (500 MHz, CDCl₃) δ 6.91–6.87 (m, 1H), 3.64 (dd, J=1.0, 2.5 Hz, 2H), 2.19 (q, J=7.5 Hz, 2H), 1.56–1.50 (m, 2H), 1.38–1.24 (m, 8H), 0.89 (t, J=7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 197.7, 191.5, 140.5, 132.8, 43.3, 31.9, 31.0, 29.6, 29.2, 28.2, 22.8, 14.3; ESIHRMS m/z calcd for $C_{12}H_{17}O_2S$ (M-1)⁻: 225.0955, found: 225.0941.

4.2.4. E-2-(Cyclopropylmethylidene)thiosuccinic anhydride (5). Rapid chromatographic purification over silica gel eluting with 10% EtOAc in hexanes afforded a colorless oil in 84% yield. IR (film) 1737 and 1699 cm^{-1} ; ¹H NMR (500 MHz, CDCl₃) δ 6.27 (dt, J=11.0, 2.5 Hz, 1H), 3.77 (d, J=2.0 Hz, 2H), 1.45-1.38 (m, 1H), 1.16-1.12 (m, 2H), 0.85-

Scheme 4. Synthesis of piperidinonyl acetamides.

0.81 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 198.2, 190.9, 145.6, 130.0, 43.6, 14.4, 10.3; ESIHRMS m/z calcd for $C_8H_7O_2S(M-1)^{-}$: 167.0172, found: 167.0166.

4.3. General procedure for the three component coupling of alkylidene thiosuccinic anhydrides with aminothiols and 2,4 dinitrobenzenesulfonamides

The aminothiol (0.5 mmol) was added to a stirred solution of the alkylidene thiosuccinic anhydride (0.5 mmol) in DMF (5 mL) at 0 °C. The reaction mixture was stirred for overnight at 0 °C before $Cs₂CO₃$ (0.6 mmol) was added, followed immediately by the sulfonamide (0.5 mmol). Stirring was continued while the reaction mixture was allowed to warm to room temperature and the subsequently for a further 1 h. The solvent was then removed under vacuum and the residue was dissolved in ethyl acetate, washed with water followed by brine, and dried over $Na₂SO₄$. Purification was then realized by silica gel column chromatography.

4.3.1. cis- and trans-N-[(Benzo[b]-5-oxo-7-phenyl-1,4-thiazepin-6yl)acetyl]piperidine (10). Chromatographic purification by eluting with 70% ethyl acetate in hexanes afforded the trans and cis isomers with yields of 60% and 12%, respectively.

trans-Isomer: pale yellow solid, mp $226-227$ °C; IR (film) 1678 and 1631 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 7.53 (d, J=7.5 Hz, 1H), 7.44 (t, J=7.7 Hz, 1H), 7.35 (d, J=7.5 Hz, 1H), 7.30–7.24 (m, 3H), 7.20 $(t, J=7.2$ Hz, 2H), 7.11 (d, J=7.5 Hz, 2H), 4.32 (d, J=12.5 Hz, 1H), 3.61 $(dt, J=2.0, 11.5 Hz, 1H), 3.46-3.39 (m, 1H), 3.36-3.24 (m, 2H), 3.18-$ 3.12 (m, 1H), 3.08 (dd, J=11.0, 16.0 Hz, 1H), 1.80 (d, J=16.0 Hz, 1H), 1.56–1.32 (m, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 174.5, 169.0, 143.4, 141.7, 136.0, 130.8, 129.2, 128.1, 126.7, 126.6, 126.3, 124.0, 56.9, 46.5, 44.6, 43.0, 33.9, 26.3, 25.6, 24.6; ESIHRMS m/z calcd for $C_{22}H_{24}N_2O_2S$ Na (M+Na)⁺: 403.1456, found: 403.1473.

 cis -Isomer: yellow solid, mp163–164 $\,^{\circ}$ C; IR (film) 1676 and 1637 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.66 (dd, J=1.5, 8.0 Hz, 1H), 7.54 (br s, 1H), 7.51 (d, J=7.0 Hz, 2H), 7.42-7.34 (m, 4H), 7.21 $(t, J=7.0$ Hz, 2H), 5.02 (d, J=7.0 Hz, 1H), 3.66–3.61 (m, 1H), 3.59–3.54 $(m, 1H)$, 3.34–3.29 $(m, 1H)$, 3.17–3.07 $(m, 2H)$, 3.53 $(dd, J=8.0, 16.5 Hz$, 1H), 2.07 (dd, J=5.5, 16.5 Hz, 1H), 1.55–1.37 (m, 6H); ¹³C NMR (125 MHz, CDCl3) d 173.8, 168.6, 141.9, 137.2, 135.3, 130.6, 129.7, 128.8, 128.6,126.7,123.4, 57.8, 46.5, 43.0, 42.1, 32.9, 26.3, 25.7, 24.7; ESIHRMS m/z calcd for C₂₂H₂₄N₂O₂SNa (M+Na)⁺: 403.1456, found: 403.1469.

4.3.2. cis- and trans-N-(2-Phenylethyl) (benzo[b]-5-oxo-7-phenyl-1,4-thiazepin-6yl)acetamide (11). Chromatographic purification eluting with 70% ethyl acetate in hexanes afforded the trans and cis isomers with yields of 63% and 13%, respectively.

trans-Isomer: pale yellow solid, mp 214–215 \degree C; IR (film) 1665 and 1642 cm^{-1} ; ¹H NMR (500 MHz, CDCl₃) δ 7.56 (d, J=8 Hz, 1H), 7.50-7.45 (m, 2H), 7.34–7.26 (m, 5H), 7.24–7.20 (m, 2H), 7.14 (d, $J=7.5$ Hz, 3H), 7.09 (d, J=6.5 Hz, 2H), 5.56 (br s, 1H), 4.27 (d, J=12 Hz, 1H), 3.55 $(dt, J=2.5, 11.5 Hz, 1H), 3.39 (q, J=6.5 Hz, 2H), 2.73-2.65 (m, 3H), 1.82$ $(dd, J=2.5, 15 Hz, 1H);$ ¹³C NMR (125 MHz, CDCl₃) δ 173.9, 170.7, 142.9, 141.1, 139.0, 136.2, 130.8, 129.3, 129.0, 128.8, 128.2, 127.1, 126.7, 126.4, 123.9, 57.1, 44.9, 40.8, 36.6, 35.8; ESIHRMS m/z calcd for $C_{25}H_{24}N_{2}O_{2}S$ Na (M+Na)⁺: 439.1456, found: 439.1457.

 cis -Isomer: pale yellow solid, mp 151-151.5 °C; IR (film) 1671 cm^{-1} ; ¹H NMR (500 MHz, CDCl₃) δ 7.66 (d, J=6.5 Hz, 1H), 7.60 (br s,1H), 7.47–7.38 (m, 3H), 7.35–7.28 (m, 4H), 7.24–7.12 (m, 6H), 5.45 $(br s, 1H)$, 4.94 (d, J=7.0 Hz, 1H), 3.57–3.52 (m, 1H), 3.47–3.34 (m, 2H), 2.72 (t, J=7.0 Hz, 2H), 2.28 (dd, J=9.0, 15.0 Hz, 1H), 1.89 (dd, J=5.0, 15.0 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 173.4, 170.7, 141.5, 138.9, 136.8, 135.4, 130.6, 129.7, 128.9, 128.8, 128.4, 127.0, 126.7, 123.4, 58.2, 42.5, 40.1, 36.2, 35.7; ESIHRMS m/z calcd for C₂₅H₂₄N₂O₂SNa $(M+Na)^+$: 439.1456, found: 439.1462.

4.3.3. cis- and trans-N-[(5-Oxo-7-phenyl-1,4-thiazepin-6yl)acetyl]piperidine (12). Chromatographic purification eluting with 4% methanol in dichloromethane afforded the trans and cis isomers with yields of 52% and 13%, respectively.

trans-Isomer: light yellow gum, IR (film) 1675 and 1614 cm^{-1} ; ¹H NMR (500 MHz, CDCl₃) δ 7.28-7.25 (m, 2H), 7.22-7.19 (m, 3H), 5.85 (br s, 1H), 4.15 (d, J=9.5 Hz, 1H), 3.66-3.54 (m, 2H), 3.52-3.43 (m, 2H), 3.33–3.28 (m, 1H), 3.14–3.05 (m, 3H), 2.96–2.85 (m, 3H), 1.50–1.18 (m, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 171.7, 171.6, 139.4, 129.7, 128.5, 126.7, 47.0, 43.8, 43.4, 43.0, 42.0, 37.3, 28.5, 25.5, 24.6; ESIHRMS m/z calcd for C₁₈H₂₄N₂O₂SNa (M+Na)⁺: 355.1456, found: 355.1445.

 cis -Isomer: light yellow gum, IR (film) 1638 cm⁻¹; ¹H NMR $(500$ MHz, CDCl₃) δ 7.37-7.27 (m, 5H), 5.95 (t, J=6.5 Hz, 1H), 4.10-4.00 (m, 2H), 3.89 (d, $J=10.0$ Hz, 1H), 3.68–3.61 (m, 1H), 3.52–3.46 (m, 1H), 3.44–3.36 (m, 1H), 3.28–3.22 (m, 1H), 3.16–3.10 (m, 1H), $2.97 - 2.89$ (m, 1H), $2.87 - 2.81$ (m, 2H), 1.88 (dd, $J = 3.0$, 16.0 Hz, 1H), 1.58–1.30 (m, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 177.0, 169.5, 140.5, 131.6, 129.3, 129.1, 128.1, 128.0, 47.3, 46.5, 46.0, 43.2, 43.0, 34.0, 32.1, 26.3, 25.6, 24.7; ESIHRMS m/z calcd for C₁₈H₂₄N₂O₂SNa (M+Na)⁺: 355.1456, found: 355.1454.

4.3.4. cis- and trans-N-(2-Phenylethyl) (5-oxo-7-phenyl-1,4-thiazepin-6yl)acetamide (13). Chromatographic purification eluting with 4% methanol in dichloromethane afforded a pale yellow gum in 67% yield as 1:4 cis/trans-mixture.

trans-Isomer: IR (film) 1666 and 1640 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.35–7.28 (m, 7H), 7.24–7.19 (m, 2H), 7.16 (d, J=7.0 Hz, 2H), 6.01 (br s, 1H), 5.52 (br s, 1H), 4.02–3.84 (m, 3H), 3.66–3.60 (m, 1H), 3.49–3.35 (m, 2H), 2.96–2.87 (m, 1H), 2.82–2.71 (m, 2H), 2.43 (dd, J=11.0, 14.7 Hz, 1H), 1.91 (dd, J=3.2, 14.7 Hz, 1H); ¹³C NMR (125 MHz, CDCl3) d 176.8, 171.5, 139.9, 139.0, 129.1, 129.0, 128.8, 128.3, 128.1, 126.6, 46.3, 43.6, 40.9, 37.5, 35.8, 32.5; ESIHRMS m/z calcd for $C_{21}H_{24}N_2O_2S$ Na (M+Na)⁺: 391.1456, found: 391.1449.

cis-Isomer: the minor cis-isomer was identified in the mixture by characteristic signals at δ 6.09 (br s, 1H), 5.63 (br s, 1H), 3.06–3.01 $(m, 1H)$, 2.53 (dd, J=8.2, 14.7 Hz, 1H), 2.09 (dd, J=4.7, 14.7 Hz, 1H) in the ¹H NMR spectrum, and by δ 175.2, 171.2, 139.3, 128.7, 128.2, 126.7, 48.0, 45.9, 40.7, 38.1, 29.8 in the ¹³C NMR spectrum.

4.3.5. cis- and trans-N-[(Benzo[b]-5-oxo-7-methyl-1,4-thiazepin-6yl)acetyl]piperidine (14). Chromatographic purification eluting with 70% ethyl acetate in hexanes afforded a pale yellow solid in 74% yield as 1:1 cis/trans-mixture.

trans-Isomer: IR (film) 1674 and 1633 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.61 (dd, J=1.5, 7.5 Hz, 1H), 7.42 (br s, 1H), 7.40–7.32 $(m, 1H)$, 7.24 $(d, J=7.0$ Hz, 1H), 7.19–7.13 $(m, 1H)$, 3.57–3.51 $(m, 1H)$, 3.47–3.38 (m, 2H), 3.33–3.27 (m, 1H), 3.16 (dd, $J=10.5$, 16 Hz, 1H), 2.92 (dt, J= 2.5 , 11 Hz, 1H), 2.24 (dd, J= 2.7 , 16.2 Hz, 1H), 1.65–1.42 (m, 6H), 1.39 (d, J=6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 175.0, 169.1, 141.5, 136.6, 135.0, 130.7, 128.3, 126.5, 125.8, 123.7, 48.3, 47.7, 46.7, 45.9, 43.2, 33.6, 26.5, 23.1, 16.9; ESIHRMS m/z calcd for $C_{17}H_{22}N_2O_2S$ Na (M+Na)⁺: 341.1300, found: 341.1291.

cis-Isomer: the cis-isomer was identified in the mixture by characteristic signals at δ 7.57 (dd, J=12.0, 7.7 Hz, 1H), 3.95–3.87 (m, 1H), 2.17 (dd, J=3.7, 16.2 Hz, 1H), 1.37 (d, J=6.5 Hz, 3H) in the ¹H NMR spectrum, and by δ 173.3, 168.7, 141.8, 123.4, 126.4, 130.2, 47.7, 43.1, 41.5, 32.5, 25.7, 24.7 in the 13C NMR spectrum.

4.3.6. cis- and trans-N-(2-Phenylethyl) (benzo[b]-5-oxo-7- methyl -1,4-thiazepin-6yl)acetamide (15). Chromatographic purification eluting with 70% ethyl acetate in hexanes afforded a pale yellow oil in 72% yield as 1:1 cis/trans-mixture.

*trans-*Isomer: IR (film) 1670 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, J=7.5 Hz, 1H), 7.57–7.30 (m, 4H), 7.25–7.14 (m, 5H), 6.15 (br s, 1H), 3.50–3.40 (m, 2H), 3.26–3.20 (m, 1H), 2.82–2.76 (m, 3H), 2.70 (dd, $J=10.5$, 14.5 Hz, 1H), 2.30 (dd, $J=2.2$, 14.2 Hz, 1H), 1.39 (d, J¼7.0 Hz, 3H); 13C NMR (125 MHz, CDCl3) d 173.1, 170.8, 141.3, 139.0, 136.7, 130.7, 129.0, 128.8, 126.8, 125.8, 123.6, 48.0, 46.6,, 41.0, 35.9, 35.6, 23.0, 16.5; ESIHRMS m/z calcd for C₂₀H₂₂N₂O₂SNa (M+Na)⁺: 377.1300, found: 377.1284.

cis-Isomer: the cis-isomer was identified in the mixture by characteristic signals at δ 7.57 (d, J=7.5 Hz, 1H), 7.08 (d, J=7.5 Hz, 1H), 5.72 (br s, 1H), 3.87 (q, J=6.5 Hz, 1H), 3.36-3.32 (m, 2H), 2.06 (dd, J=3.5, 14.7 Hz, 1H), 1.31 (d, J=6.5 Hz, 3H) in the ¹H NMR spectrum, and by δ 175.1, 170.9, 141.1, 139.2, 135.1, 130.2, 128.1, 126.7, 123.4, 48.8, 42.3, 40.8, 36.5, 35.87 in the ¹³C NMR spectrum.

4.3.7. cis- and trans-N-[(5-Oxo-7-methyl-1,4-thiazepin-6yl)acetyllpiperidine (16). Chromatographic purification eluting with 4% methanol in dichloromethane afforded a pale yellow oil in 58% yield as 1:1 cis/trans-mixture.

*trans-*Isomer: IR (film) 1663 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 6.00 (br s, 1H), 3.93-3.97(m, 1H), 3.60-3.49 (m, 5H), 3.40 (dt, $J=3.5, 9.7$ Hz, 1H), $3.11-3.05$ (m, 1H), 2.98 (dd, $J=11.7, 14.2$ Hz, 1H), 2.54 (dd, $J=11.5$, 14.5 Hz, 1H), 2.37 (dd, $J=3.2$, 16.2 Hz, 1H), 1.70–1.51 (m, 6H), 1.38 (d, J=7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 176.2, 169.2, 48.5, 46.7, 45.7, 43.1, 36.0, 33.2, 26.6, 25.8, 24.8, 20.3, 16.7; ESIHRMS m/z calcd for C₁₃H₂₂N₂O₂SNa (M+Na)⁺: 293.1300, found: 293.1284.

cis-Isomer: the cis-isomer was identified in the mixture by characteristic signals at δ 5.87 (br s, 1H), 4.03 (dd, J=5.5, 8.0 Hz, 1H), 3.80–3.73 (m, 1H), 3.48–3.41 (m, 5H), 2.88 (dt, $J=4.5$, 14.5 Hz, 1H), 2.83-2.74 (m, 3H), 2.24 (dd, J=5.5, 16.0 Hz, 1H), 1.37 (d, *J*=7.0 Hz, 3H) in the ¹H NMR spectrum, and by δ 176.9, 169.4, 48.3, 46.8, 43.2, 42.2, 35.9, 34.7, 27.3, 26.5, 25.7 in the 13C NMR spectrum.

4.3.8. cis- and trans-N-(2-Phenylethyl) (5-oxo-7-methyl-1,4-thiazepin-6yl)acetamide (17). Chromatographic purification eluting with 4% methanol in dichloromethane afforded a pale yellow oil in 62% yield as 1:1 cis/trans-mixture.

trans-Isomer: IR (film) 1649 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 7.31 (t, J=7.5 Hz, 2H), 7.24–7.19 (m, 3H), 6.09 (br s, 1H), 5.96 (br s, 1H), 3.83–3.77 (m, 1H), 3.66–3.59 (m, 1H), 3.57–3.44 (m, 3H), 2.87 $(dt, J=4.2, 19.7 Hz, 1H), 2.82-2.77 (m, 4H), 2.52 (dd, J=5.5, 15.0 Hz,$ 1H), 2.16 (dd, J=4.7, 14.2 Hz, 1H), 1.33 (d, J=7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl3) d 176.0, 171.2, 139.2, 129.0, 126.7, 49.2, 45.8, 42.2, 40.9, 38.6, 37.0, 36.2, 26.9, 20.3, 16.5; ESIHRMS m/z calcd for $C_{16}H_{22}N_2O_2S$ Na (M+Na)⁺: 329.1300, found: 329.1302.

cis-Isomer: the cis-isomer was identified in the mixture by characteristic signals at δ 6.07 (br s, 1H), 5.94-5.90 (m, 1H), 3.89 (dd, $J=4.5$, 9.5 Hz, 1H), 3.27-3.21 (m, 1H), 2.94 (dd, $J=10.7$, 15.0 Hz, 1H), 2.75-2.66 (m, 2H), 2.38 (dd, J=3.7, 14.2 Hz, 1H),1.38 (d, J=7.0 Hz, 3H) in the ¹H NMR spectrum, and by δ 176.5, 171.3, 139.1, 128.8, 126.6, 49.1, 40.8, 36.3, 35.9, 28.9 in the ¹³C NMR spectrum.

4.3.9. cis- and trans-N-[(Benzo[b]-5-oxo-7-octanyl-1,4-thiazepin-6yl)acetyl]piperidine (18). Chromatographic purification eluting with 70% ethyl acetate in hexanes afforded a pale yellow oil in 67% yield as 2:5 cis/trans-mixture.

trans-Isomer: IR (film) 1677 and 1641 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.57 (t, J=8.0 Hz, 1H), 7.51 (br s, 1H), 7.34–7.31 (m, 1H), 7.23 (d, J¼8.0 Hz, 1H), 7.17–7.12 (m, 1H), 3.53–3.36 (m, 4H), 3.19–3.09 $(m, 1H)$, 3.00 (dt, J=2.0, 11.0 Hz, 1H), 2.26 (d, J=15.5 Hz, 1H), 1.84– 1.70 (m, 1H), 1.50–1.56 (m, 4H), 1.56–1.40 (m, 5H), 1.40–1.14 (m, 9H), 0.92–0.83 (m, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 175.3, 169.2, 141.8, 136.6, 130.6, 126.4, 126.3, 123.6, 54.2, 46.7, 44.6, 43.2, 43.1, 34.9, 33.5, 32.0, 29.8, 29.4,, 26.5, 25.7, 24.8, 22.9, 14.3; ESIHRMS m/z calcd for $C_{23}H_{34}N_2O_2S$ Na (M+Na)⁺: 425.2239, found: 425.2249.

cis-Isomer: the cis-isomer was identified in the mixture by characteristic signals at δ 7.65 (br s, 1H), 7.32 (t, J=7.5 Hz, 3H), 7.05 (d, J=7.5 Hz, 1H), 3.78-3.74 (m, 1H), 2.16 (dd, J=3.2, 16.2 Hz, 1H) in the ¹H NMR spectrum, and by δ 173.6, 168.8, 135.1, 130.0, 128.1, 123.4, 53.6, 41.4, 32.7, 27.7, 26.4, 24.7 in the 13C NMR spectrum.

4.3.10. cis- and trans-N-(2-Phenylethyl) (benzo[b]-5-oxo-7-octanyl -1,4-thiazepin-6yl)acetamide (19). Chromatographic purification eluting with 70% ethyl acetate in hexanes afforded a pale yellow oil in 61% yield as 2:5 cis/trans-mixture.

trans-Isomer: IR (film) 1672 cm^{-1} ; ¹H NMR (500 MHz, CDCl₃) δ 7.61–7.54 (m, 2H), 7.38 (dt, J=1.5, 7.5 Hz, 1H), 7.35–7.30 (m, 3H), 7.25–7.14 (m, 4H), 6.18 (br s, 1H), 3.54–3.41 (m, 2H), 3.06 $(t, J=10.0$ Hz, 1H), 2.88–2.70 (m, 4H), 2.32 (dd, $J=2.5$, 14.0 Hz, 1H), 1.67–1.52 (m, 2H), 1.45–1.20 (m, 10H), 0.92–0.88 (m, 3H); 13C NMR (125 MHz, CDCl3) d 175.2, 171.1, 141.4, 139.2, 136.7, 135.3, 130.6, 129.0, 128.5, 126.8, 126.7, 123.6, 54.6, 45.3, 40.9, 36.6, 35.8, 34.7, 32.0, 29.7, 29.4, 26.4, 22.9, 14.3; ESIHRMS m/z calcd for $C_{26}H_{34}N_2O_2S$ Na (M+Na)⁺: 461.2239, found: 461.2261.

cis-Isomer: the cis-isomer was identified in the mixture by characteristic signals at δ 7.66 (br s, 1H), 7.07 (d, J=8.0 Hz, 1H), 5.76 (br s, 1H), 3.77–3.72 (m, 1H), 3.40–3.35 (m, 1H), 2.06 (dd, $J=3.7$, 14.2 Hz, 1H), 1.78-1.72 (m, 1H) in the 1 H NMR spectrum, and by d 173.2, 170.8, 141.2, 139.0, 130.1, 126.4, 123.3, 53.8, 42.2, 31.7, 27.5 in the 13 C NMR spectrum.

4.3.11. cis- and trans-N-[(Benzo[b]-5-oxo-7-cyclopropyl-1,4-thiazepin-6yl)acetyl]piperidine (20). Chromatographic purification eluting with 70% ethyl acetate in hexanes afforded a pale yellow oil in 57% yield as 2:3 cis/trans-mixture.

trans-Isomer: IR (film) 1677 and 1638 cm⁻¹; ¹H NMR $(500 \text{ MHz}, \text{CDCl}_3)$ δ 7.55 (dd, J=1.0, 7.5 Hz, 1H), 7.38–7.31 (m, 2H), 7.24–7.12 (m, 2H), 3.58–3.35 (m, 5H), 3.26 (dd, J=3.0, 11.5 Hz, 1H), 3.20–3.15 (m, 1H), 3.09 (dd, J=11.0, 16.0 Hz, 1H), 2.46 (dd, J=5.0, 16.5 Hz, 1H), 1.52–1.40 (m, 6H), 0.76–0.60 (m, 2H), 0.35–0.25 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 173.9, 169.0, 141.6, 135.1, 130.1, 126.6, 123.5, 59.0, 45.3, 42.4, 32.8, 25.7, 24.7, 18.4, 12.5, 7.1, 3.9, ESIHRMS m/z calcd for C₁₉H₂₄N₂O₂SNa (M+Na)⁺: 367.1456, found: 367.1450.

cis-Isomer: the minor cis-isomer was identified in the mixture by characteristic signals at δ 7.63 (dd, J=1.0, 4.5 Hz, 1H), 2.73–2.67 (m, 1H), 0.53–0.48 (m, 2H) in the ¹H NMR spectrum, and by δ 174.8,

169.4, 141.3, 136.1, 130.4, 128.7, 126.4, 123.4, 59.4, 46.7, 43.1, 33.5, 26.5, 9.0, 3.1 in the 13 C NMR spectrum.

4.3.12. cis- and trans-N-(2-Phenylethyl) (benzo[b]-5-oxo-7-cyclopropyl-1,4-thiazepin-6yl)acetamide (21). Chromatographic purification eluting with 70% ethyl acetate in hexanes afforded a pale yellow oil in 60% yield as 2:3 cis/trans-mixture.

*trans-*Isomer, IR (film) 1671 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 7.65 (dd, J=1.0, 7.5 Hz, 1H), 7.40–7.30 (m, 4H), 7.25–7.15 (m, 5H), 5.93 (br s, 1H), 3.53–3.45 (m, 2H), 3.22–3.17 (m, 1H), 2.79 $(q, J=7.0$ Hz, 2H), 2.70 $(d, J=7.0$ Hz, 2H), 1.04–0.97 (m, 1H), 0.77–0.59 (m, 3H), 0.32–0.22 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 174.6, 171.3, 140.8, 139.2, 136.2, 130.4, 129.0, 128.9, 128.8 126.4, 123.4, 59.5, 45.9, 40.9, 36.7, 35.9, 31.5, 18.2, 9.0, 3.8; ESIHRMS m/z calcd for $C_{22}H_{24}N_2O_2S$ Na (M+Na)⁺: 403.1456, found: 403.1472.

cis-Isomer: the minor cis-isomer was identified in the mixture by characteristic signals at δ 7.56 (dd, J=1.2, 7.7 Hz, 1H), 7.09 $(d, J=7.5 \text{ Hz}, 1\text{ H}), 5.72 \text{ (br s, 1H)}, 3.42-3.38 \text{ (m, 1H)}, 3.11 \text{ (dd, } J=5.5,$ 10.5 Hz, 1H), 3.00 (dd, J=9.7, 14.7 Hz, 1H), 2.64 (dd, J=9.5, 11.5 Hz, 1H), 2.26 (dd, J=4.5, 14.5 Hz, 1H), 1.19-1.14 (m, 1H), 0.92-0.86 (m, 1H), 0.51–0.46(m, 1H) in the 1 H NMR spectrum, and by δ 173.5, 171.1, 141.2, 135.1, 130.2, 128.5, 126.8, 126.7, 123.5, 59.4, 43.1, 35.9, 25.9, 12.4, 7.2, 3.0 in the ¹³C NMR spectrum.

4.3.13. 2-[3-(tert-Butyloxycarbonylamino)-2-methylpropyl]succinic thioanhydride (23). Tris(trimethylsilyl)silane (373 mg, 1.5 mmol) and AIBN (33 mg, 0.2 mmol) in dry degassed toluene (5 mL) were added drop-wise to a stirred mixture of 22 (407 mg, 1.5 mmol) and 3 (142 mg, 1 mmol) in dry degassed toluene (10 mL) at 90 \degree C over 5 h by syringe pump under a N_2 atmosphere. When the addition was complete, the reaction mixture was allowed to stir for an additional 1 h at 90 °C before it was cooled to room temperature and the solvent was removed under vacuum. Rapid chromatographic purification over silica gel, pre-washed with acetone followed by hexanes, eluting with 30% ethyl acetate in hexanes afforded a colorless oil in 52% yield as a 3:1 inseparable anti/syn diastereomeric mixture.

anti-Isomer: IR (film) 1708 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 4.58 (br s, 1H), 3.55–3.02 (m, 4H), 2.88 (dd, J=4.0, 6.5 Hz, 1H), 2.26–2.18 (m, 1H), 1.66–1.58 (m, 1H), 1.54–1.41 (m, 10H), 1.04 (d, J=7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 203.0, 199.9, 156.2, 79.6, 57.4, 44.4, 38.6, 35.7, 33.3, 28.6, 17.2; ESIHRMS m/z calcd for $C_{13}H_{21}NO_4$ SNa (M+Na)⁺: 310.1089, found: 310.1082.

syn-Isomer: the minor syn-isomer was identified in the mixture by characteristic signals at δ 2.84 (dd, J=4.0, 6.5 Hz, 1H), 2.44– 2.36 (m, 1H), 0.96 (d, J=7.0 Hz, 3H) in the ¹H NMR spectrum, and by δ 199.8, 56.9, 42.3, 38.3, 32.6, 31.5, 14.7 in the ¹³C NMR spectrum.

4.4. General procedure for piperidinone synthesis from the radical adduct 23

To a stirred solution of 23 (145 mg, 0.5 mmol) in dichloromethane (20 mL) at 0° C, trifluoroacetic acid (5 mL) was added drop-wise. Stirring was maintained for 1 h before the acid was removed by azeotropic distillation with toluene $(5 \text{ mL} \times 3)$, after which the residue was dried under vacuum. The residue was dissolved in DMF (20 mL) and cooled down to 0° C before 2,4,6-collidine (0.090 mg, 0.75 mmol) was added drop-wise and the reaction mixture was stirred for 1 h at 0° C. The resulting reaction mixture was maintained at 0° C, and Cs_2CO_3 (0.195 mg, 0.6 mmol) followed by the sulfonamide (0.6 mmol) was added. The reaction mixture was warmed to room temperature and stirred for 1.5 h before the solvent was removed under vacuum and the residue was dissolved in EtOAc (30 mL). The organic layer was washed successively with water and brine, dried over $Na₂SO₄$, concentrated, and purified by silica gel column chromatography.

4.4.1. N-(2-Phenylethyl) (4-methyl-2-oxopiperidin-3-yl)acetamide (24). Chromatographic purification eluting with 4% methanol in dichloromethane afforded a pale yellow solid in 78% yield as a 3:1 cis/trans-mixture.

cis-Isomer: IR (film) 1650 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.30 (t, J=7.5 Hz, 2H), 7.24-7.19 (m, 3H), 6.86 (br s, 1H), 5.78 (br s, 1H), 3.54–3.45 (m, 2H), 3.38 (dt, J=5.5, 11.5 Hz, 1H), 3.32–3.23 (m, 1H), 2.84–2.79 (m, 2H), 2.77–2.70 (m, 1H), 2.28–2.21 (m, 1H), 2.09 (dd, J¼4.5, 14.5 Hz, 1H), 2.01–1.94 (m, 1H), 1.88–1.78 (m, 2H), 0.93 (d, $=$ 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 174.7, 172.6, 139.4, 129.0, 126.5, 43.4, 40.9, 39.1, 36.4, 35.9, 30.6, 28.9, 13.9; ESIHRMS m/z calcd for $C_{16}H_{22}N_2O_2Na$ $(M+Na)^+$: 297.1579, found: 297.1591.

trans-Isomer: the minor trans-isomer was identified in the mixture by characteristic signals at δ 6.50 (br s, 1H), 5.85 (br s, 1H),2.50 (dd, J=6.0, 14.5 Hz, 1H), 2.18–2.14 (m, 1H), 1.72–1.66 (m, 1H) 1.1 (d, J=6.5 Hz, 3H) in the ¹H NMR spectrum, and by δ 174.4, 171.9, 139.3, 129.0, 128.7, 46.6, 41.3, 40.7, 36.0, 35.8, 31.9, 30.7, 20.4 in the 13C NMR spectrum.

4.4.2. N-[(4-Methyl-2-oxopiperidin-3-yl)acetyl]piperidine (25). Chromatographic purification eluting with 4% methanol in dichloromethane afforded the cis- and trans-isomers with yields of 56% and 19%, respectively.

 cis -Isomer: light yellow gum, IR (film) 1635 cm⁻¹; ¹H NMR (500 MHz, CDCl3) d 5.63 (br s, 1H), 3.64–3.59 (m, 1H), 3.56–3.39 (m, 4H), 3.34–3.29 (m, 1H), 3.14–3.09 (m, 2H), 2.43–2.37 (m, 1H), 2.27 (dd, J=9.7, 17.2 Hz, 1H), 2.09-2.03 (m, 1H), 1.75-1.52 (m, 7H), 0.96 (d, $J=7.0$ Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 174.1, 170.0, 46.8, 43.1, 42.9, 38.9, 30.6, 29.3, 28.5, 26.7, 25.8, 24.9, 14.0; ESIHRMS m/z calcd for C₁₃H₂₂N₂O₂Na (M+Na)⁺: 261.1579, found: 261.1561.

trans-Isomer: light yellow gum, IR (film) 1646 cm^{-1} ; ¹H NMR $(500 \text{ MHz}, \text{CDCl}_3)$ δ 5.72 (br s, 1H), 3.59–3.40 (m, 5H), 3.32–3.26 $(m, 1H)$, 2.94 (dd, J=5.5, 16.5 Hz, 1H), 2.70 (dd, J=3.2, 16.2 Hz, 1H), 2.26–2.20 (m, 1H), 2.17–2.12 (m, 1H), 1.85 (dd, $J=3.2$, 13.2 Hz, 1H), 1.68–1.50 (m, 7H), 1.02 (d, J=7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) d 174.6, 169.6, 46.7, 45.6, 43.0, 41.2, 37.9, 32.1, 31.5, 31.1, 29.9, 26.6, 25.8, 24.8, 20.6; ESIHRMS m/z calcd for $C_{13}H_{22}N_{2}O_{2}Na$ (M+Na)⁺: 261.1579, found: 261.1564.

Acknowledgements

We thank the NIH (GM57335 and GM62160) for support of this work.

Supplementary data

Supplementary data associated with this article can be found in online version at doi:10.1016/j.tet.2010.04.002.

References and notes

- 1. Niyomura, O.; Kato, S. Top. Curr. Chem. 2005, 251, 1–12.
- 2. Kato, S.; Kawahara, Y.; Kageyama, H.; Yamada, R.; Niyomura, O.; Murai, T.; Kanda, T. J. Am. Chem. Soc. 1996, 118, 1262–1267.
- 3. Hadad, C. M.; Rablen, P. R.; Wiberg, K. B. J. Org. Chem. 1998, 63, 8668–8681.
- 4. Bauer, W.; Kühlein, K. Methoden der Organischen Chemie In Carbonsäure und
- Carbonsäure Derivate, 4th ed.; Falbe, J., Ed.; Thieme: Stuttgart, 1985; pp 832-890. 5. Kato, S.; Murai, T. In The Chemistry of Acid Derivatives; Patai, S., Ed.; Wiley: Chichester, UK, 1992; pp 803–847.
- 6. Scheithauer, S.; Mayer, R. Top. Sulfur Chem. 1979, 4, 1–373.
- 7. Lu, W.; Qasim, M. A.; Kent, S. B. H. J. Am. Chem. Soc. 1996, 118, 8518–8523.
- 8. Crich, D.; Sharma, I. Angew. Chem., Int. Ed. 2009, 48, 2355–2358.
- 9. Talan, R. S.; Sanki, A. K.; Sucheck, S. J. Carbohydr. Res. 2009, 344, 2048–2050.
- 10. Messeri, T.; Sternbach, D. D.; Tomkinson, N. C. O. Tetrahedron Lett. 1998, 39, 1673–1676.
- 11. Messeri, T.; Sternbach, D. D.; Tomkinson, N. C. O. Tetrahedron Lett. 1998, 39, 1669–1672.
- 12. Crich, D.; Sasaki, K. Org. Lett. 2009, 11, 3514–3517.
- 13. Kricheldorf, H. R.; Leppert, E. Makromol. Chem. 1973, 167, 47–68.
- 14. Stockdill, J. L.; Wua, X.; Danishefsky, S. J. Tetrahedron Lett. 2009, 50, 5152–5155.
- 15. Wua, X.; Li, X.; Danishefsky, S. J. Tetrahedron Lett. 2009, 50, 1523–1525.
- 16. Yuan, Y.; Zhu, J.; Li, X.; Wua, X.; Danishefsky, S. J. Tetrahedron Lett. 2009, 50, 2329–2333.
- 17. Bao, Y.; Li, X.; Danishefsky, S. J. J. Am. Chem. Soc. 2009, 131, 12924-12926.
- 18. Blake, J. Int. J. Pept. Protein Res. 1981, 17, 273–274.
- 19. Yamashiro, D.; Blake, J. F. Int. J. Pept. Protein Chem. 1981, 18, 383–392.
- 20. Mitin, Y. V.; Zapevalova, N. P. Int. J. Pept. Protein Chem. 1990, 35, 352–356.
- 21. Høeg-Jensen, T.; Olsen, C. E.; Holm, A. J. Org. Chem. 1994, 59, 1257–1263.
- 22. Kolakowski, R. V.; Shangguan, N.; Sauers, R. R.; Williams, L. J. J. Am. Chem. Soc. 2006, 128, 5695–5702.
- 23. Kolakowski, R. V.; Shangguan, N.; Williams, L. J. Tetrahedron Lett. 2006, 47, 1163–1166.
- 24. Shangguan, N.; Katukojvala, S.; Greenberg, R.; Williams, L. J. J. Am. Chem. Soc. 2003, 125, 7754–7755.
- 25. Barlett, K. N.; Kolakowski, R. V.; Katukojvala, S.; Williams, L. J. Org. Lett. 2006, 8, 823–826.
- 26. Merkx, R.; Brouwer, A. R.; Rijkers, D. T. S.; Liskamp, R. M. J. Org. Lett. 2005, 7, 1125–1128.
- 27. Merkx, R.; van Haren, M. J.; Rijkers, D. T. S.; Liskamp, R. M. J. J. Org. Chem. 2007, 72, 4574–4577.
- 28. Rakotomanomana, N.; Lacombe, J.-M.; Pavia, A. Carbohydr. Res.1990,197, 318–323.
- 29. Rosen, T.; Lico, I. M.; Chu, D. T. W. J. Org. Chem. 1988, 53, 1580–1582.
- 30. McKervey, M. A.; O'Sullivan, M. B.; Myers, P. L.; Green, R. H. Chem. Commun. 1993, 94–96.
- 31. Crich, D. In Organosulfur Chemistry: Synthetic Aspects; Page, P., Ed.; Academic: London, 1995; pp 49–88.
- 32. Stacey, F. W.; Harris, J. F. Org. React. 1963, 13, 150–376.
- 33. Weiewer, M.; Chaminade, X.; Bayon, J. C.; Dunach, E. Eur. J. Org. Chem. 2007, 2464–2469.
- 34. Weiwer, M.; Dunach, E. Tetrahedron Lett. 2005, 47, 287–289.
- 35. Crich, D.; Sana, K. J. Org. Chem. 2009, 74, 7383–7388.
- 36. Crich, D.; Sana, K.; Guo, S. Org. Lett. 2007, 9, 4423–4426.
- 37. Rao, Y.; Li, X.; Nagorny, P.; Hayashida, J.; Danishefsky, S. J. Tetrahedron Lett. 2009, 50, 6684–6686.
- 38. Crich, D.; Rahaman, M. Y. J. Org. Chem. 2009, 74, 6792–6796.
- 39. Crich, D.; Sasaki, K.; Rahaman, M. Y.; Bowers, A. A. J. Org. Chem. 2009, 74, 3886–3893.
- 40. Crich, D.; Bowers, A. A. Org. Lett. 2007, 9, 5323–5325.
- 41. Mapp, A. K.; Ansari, A. Z.; Ptashne, M.; Dervan, P. B. Proc. Natl. Acad. Sci. U.S.A. 2000, 97, 3930–3935.
- 42. Mapp, A. K.; Dervan, P. B. Tetrahedron Lett. 2000, 41, 9451–9454.
- 43. Crich, D.; Sana, K. J. Org. Chem. 2009, 74, 3389–3393.
- 44. Aksnes, G. Acta Chem. Scand. 1961, 15, 692–694.
- 45. Bacaloglu, R.; Blasko, A.; Bunton, C. A.; Cerichelli, G.; Castaneda, F.; Rivera, E. J. Chem. Soc., Perkin 2 1995, 965–972.
- 46. Balasubramaniyan, V.; Tongare, D. B.; Gosavi, S. S.; Babar, S. M. Proc. Ind. Acad. Sci., Chem. Sci. 1993, 105, 265–271.
- 47. Bourdel, E.; Doulut, S.; Jarretou, G.; Labbe-Jullie, C.; Fehrentz, J.-A.; Doumbia, O.; Kitabgi, P.; Martinez, J. Int. J. Pept. Protein Res. 1996, 48, 148–155.
- 48. Cunha, S.; Kascheres, A. J. Brazilian Chem. Soc. 2000, 11, 525–529.
- 49. Doulut, S.; Dubuc, I.; Rodriguez, M.; Vecchini, F.; Fulcrand, H.; Barelli, H.; Checler, F.; Bourdel, E.; Aumelas, A.; Lallement, J. C.; Kitabgi, P.; Costentin, J.; Martinez, J. J. Med. Chem. 1993, 36, 1369–1379.
- 50. Hudson, R. F.; Chopard, P. A. Helv. Chim. Acta 1963, 46, 2178–2185.
- 51. Osuch, C.; Franz, J. E.; Zienty, F. B. J. Org. Chem. 1964, 29, 3721–3722.
- 52. Kates, M. J.; Schauble, J. H. J. Org. Chem. 1995, 60, 6676–6677.
- 53. Kates, M. J.; Schauble, J. H. J. Heterocycl. Chem. 1995, 32, 971–978.
- 54. Structures and stereochemistries were assigned based on a combination of one and two dimensional NMR experiments.
- 55. McGarvey, G. J.; Williams, J. M. J. Am. Chem. Soc. 1985, 107, 1435–1438.
- 56. Yamamoto, Y.; Yamada, J.-i.; Uyehara, T. J. Am. Chem. Soc. 1987, 109, 5820–5822.
- 57. Fleming, I.; Hill, J. H. M.; Parker, D.; Waterson, D. J. Chem. Soc., Chem. Commun. 1985, 318–319.
- 58. Crump, R. A. N. C.; Fleming, I.; Hill, J. H. M.; Parker, D.; Reddy, N. L.; Waterson, D. J. Chem. Soc., Perkin Trans. 1 1992, 3277–3294.
- 59. Paddon-Row, M. N.; Rondan, N. G.; Houk, K. N. J. Am. Chem. Soc. 1982, 104, 7162–7166.
- 60. Zimmerman, H. E. Acc. Chem. Res. 1987, 20, 263–268.
- 61. Davies, H. M. L.; Hodges, L. M.; Gregg, T. M. J. Org. Chem. 2001, 66, 7898–7902.
- 62. Mohrig, J. R.; Rosenberg, R. E.; Apostol, J. W.; Bastienaansen, M.; Evans, J. W.; Franklin, S. J.; Frisbie, C. D.; Fu, S. S.; Hamm, M. L.; Hirose, C. B.; Hunstad, D. A.; James, T. L.; King, R. W.; Larson, C. J.; Latham, H. A.; Owen, D. A.; Stein, K. A.; Warnet, R. J. Am. Chem. Soc. 1997, 119, 479–486.
- 63. Banfi, L.; Guanti, G. Tetrahedron: Asymmetry 1999, 10, 439–447.
- 64. Davies, S. G.; Garner, A. C.; Long, M. J. C.; Morrison, R. M.; Roberts, P. M.; Savory, E. D.; Smith, A. D.; Sweet, M. J.; Withey, J. M. Org. Biomol. Chem. 2005, 3, 2762–2775.
- 65. Karabatsos, G. J.; Fenoglio, D. J. Top. Stereochem. 1970, 5, 167–203.
- 66. Hoffmann, R. W. Chem. Rev. 1989, 89, 1841–1860.
- 67. This model assumes kinetic selectivity. We were unable to isolate the intermediate adducts before rearrangement to the thiazepinone system and so could not test for equilibration at that level. The cis- and trans- thiazepinones were somewhat resistant to equilibration under mild acid and basic conditions suggesting that at least equilibration did not occur after rearrangement.
- 68. Chatgilialoglu, C. Acc. Chem. Res. 1992, 25, 188–194.
- 69. Giese, B.; Damm, W.; Wetterlich, F.; Zeitz, H.-G. Tetrahedron Lett. 1992, 1863–1866.
- 70. Hart, D. J.; Huang, H.-C. Tetrahedron Lett. 1985, 26, 3749–3752.
- 71. Kopping, B.; Chatgilialoglu, C.; Zehnder, M.; Giese, B. J. Org. Chem. 1992, 57, 3994–4000.
- 72. Durkin, K. A.; Liotta, D. C.; Rancourt, J.; Lavellée, J.-F.; Boisvert, L.; Guindon, Y. J. Am. Chem. Soc. 1992, 114, 4912–4914.
- 73. Erdmann, P.; Schafer, P.; Springer, R.; Zeitz, H.-G.; Giese, B. Helv. Chim. Acta 1992, 75, 638–644.
- 74. Curran, D. P.; Porter, N. A.; Giese, B. Stereochemistry of Radical Reactions; VCH: Weinheim, 1996
- 75. Equilibration of the isomers of 25 with sodium methoxide in hot methanol resulted in an equilibrium mixture favoring the trans-isomer indicating the kinetic nature of the products observed.